

ASCORBATE INHIBITS THE ENDOTHELIUM-DEPENDENT HYPERPOLARIZATION MEDIATED RELAXATION IN RESISTANCE ARTERIES

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Abstract

The main path of endothelium-dependent relaxation (EDR) in resistance arteries is independent of nitric oxide synthase and cyclooxygenase and is mediated by endothelium-dependent hyperpolarisation (EDH). We tested the effect of ascorbate upon this path in isolated small mesenteric arteries from the rat. EDR was induced by carbachol 10^{-5} M, in rings contracted by phenylephrine 10^{-5} M or by prostaglandin $F_{2\alpha}$ 10^{-5} M, in the presence of L-N(γ)-Nitro arginine methyl ester (L-NAME) 10^{-4} M and indomethacin 10^{-5} M. EDR proved to be reduced by ascorbate concentrations ranging between 10^{-5} M - 10^{-3} M ($n = 6$; $p < 0.01$). We discuss this new effect in the context of the relations between ascorbate and endothelial function/dysfunction.

Rezumat

Calea principală de relaxare endoteliiu-dependență (RED) în artere de rezistență nu depinde de nitric oxid sintază și ciclooxigenază și este mediată de hiperpolarizarea endoteliiu-dependență (HED). Am testat efectul ascorbatului asupra acestei căi, în artere mezenterice mici izolate de la șobolan. RED a fost indusă de carbacol 10^{-5} M, în inele arteriale contractate cu fenilefrină 10^{-5} M sau cu prostaglandină $F_{2\alpha}$ 10^{-5} M, în prezența esterului metilic al metilic L-N(γ)-Nitro-argininei (L-NAME) 10^{-4} M și a indometacinului 10^{-5} M. RED este redusă de ascorbat 10^{-5} M - 10^{-3} M ($n = 6$; $p < 0,01$). Studiul evaluează acest nou efect în contextul relațiilor dintre ascorbat și funcția/disfuncția endotelială.

Keywords: vitamin C, small arteries, EDH-mediated relaxation, oxidative stress

Introduction

Ascorbate (ASC) could be beneficial against endothelial dysfunction [23]. The experimental and clinical data are still controversial, as presented by the most recent reviews [26]. A review has described the benefit of ASC against endothelial dysfunction, but only for high daily doses [3]. Others have found no cardiovascular benefit from ascorbate supplementation [8]. A recent meta-analysis (one main trial with almost 15000 cases) found no cardiovascular benefit from ASC [2]. The essential NOS co-factor tetrahydrobiopterin (BH4) is key to ASC interaction with NO-mediated EDR; ASC benefits *via* BH4 protection with recovery from low NOS activity [4, 10, 11].

Resistance arteries and arterioles together are the main contributor to peripheral resistance. Endothelium-dependent relaxation (EDR) of vascular smooth muscle (VSM) is known to be mediated by nitric oxide (NO). Still, the major EDR mechanism in resistance arteries is independent of NO-synthase (NOS) and cyclooxygenase (COX), and it is mediated by endothelium-dependent hyperpolarisation (EDH) [13]. ASC produces direct VSM relaxation and EDR and modulates NO relaxing potency and guanylate cyclase (GC) [10, 23]. EDH-

mediated EDR relies on endothelial Ca^{2+} -activated K^+ channels (K_{Ca}) [5-7, 28], with conductances that are small (sK) or intermediate (iK). EDH-mediated EDR is: a counter-regulator of sympathetic vasoconstriction [18]; important in flow autoregulation, myogenic response and vasomotion [13]; compensatory in endothelial dysfunction, but also involved in its pathogenesis [12, 15-17].

Very few studies on ASC and EDH-mediated EDR have used isolated rabbit iliac artery [14], isolated perfused bovine ciliary bed [24, 25], and isolated perfused rat mesenteric arterial bed [25]. Given the sustained broad interest in EDH-mediated EDR [13] and our sustained interest in endothelial function and dysfunction [9, 19, 30-32] and in various aspects of oxidative stress [20, 22, 34, 35], we decided to test the effect of ASC upon EDH-mediated EDR in isolated small mesenteric arteries. One previous study has examined EDH-mediated EDR in isolated small mesenteric arteries [33], but ours differs in some aspects, and we discuss their relevance.

Materials and Methods

Chemicals

The salts used for the physiological saline solution (PSS) containing (mM): 119 NaCl; 4.7 KCl; 2.5 CaCl₂; 1.2 MgSO₄; 25 NaHCO₃; 1.18 KH₂PO₄; 5.5 glucose were of analytical grade (Merck, USA), and all drugs were of research grade (Sigma, USA).

Animals and procedures

Male adult Wistar rats (~ 200 g) were euthanised. Jejunal loops were excised and placed in PSS at ~ 5°C, second-order mesenteric arteries were dissected and rings 1 mm wide were cut out and mounted in wire myographs, then stretched to 1 g passive tension. The PSS was then kept at 37°C and aerated with 95% O₂ + 5% CO₂ (pH 7.2 - 7.4). After 30 min. equilibration, each ring was checked twice for complete EDR by carbachol 10⁻⁵ M upon contraction by phenylephrine 10⁻⁵ M (PHE). Then, after 30 min. re-equilibration, the contraction was induced randomly by either PHE 10⁻⁵ M or prostaglandin F_{2α} 10⁻⁵ M (PGF), and global EDR or EDH-mediated EDR was tested; first in the absence of ASC and then in the presence of ASC (10⁻⁵ M, or 10⁻⁴ M, or 10⁻³ M; actual tests) or again in its absence (control). EDH-mediated EDR was obtained in the presence of indomethacin 10⁻⁵ M to inhibit COX and N(γ)-nitro-L-arginine methyl ester (L-NAME) 10⁻⁴ M to inhibit NOS. Only one of these 16 conditions (12 test and 4 control) was randomly applied to any of the rings tested. Time-match paired rings confirmed the 30 min. stability of contraction (PHE and PGF; without carbachol).

These tests in arterial rings in isometric conditions were done with absolutely no PSS flow, even during drug administration (concentrated solution of the drug added to the organ bath; < 1/10 v/v) or removal (repeated replacement of the bath content with fresh PSS with no drugs). This study was approved by the Research Ethics Commission of the host institution. All procedures comply with guidelines from Act on Animal Experimentation and Animal Health and Welfare from Romania; Universities Federation for Animal Welfare; Directive 2010/63/EU of the European Parliament, and of the Council of 22 September 2010.

Statistical analysis

EDR in the presence of ASC was expressed as % of the control value previously obtained in the absence of ASC in each ring. Results are presented as mean ± SD (n = 6 in all series). Data were assessed using two-way ANOVA and t-test. Statistical significance was considered for p < 0.05.

Results and Discussion

ASC 10⁻⁵ - 10⁻³ M inhibits EDH-mediated EDR induced by carbachol 10⁻⁵ M (without effect on the global EDR) in rat isolated resistance mesenteric arteries when they are precontracted by any of the two agents used (Table I) (dose-dependent ASC effect; IC₅₀ 0.078 mM with PHE contraction; IC₅₀ 0.114 mM with PGF contraction). This effect is not detectable in the main mesenteric artery (data not shown).

Table I

Ascorbate effects upon endothelium-dependent relaxation (EDR) in rat isolated resistance mesenteric arteries

| contraction | PHE | PHE | PGF | PGF |
|------------------------|------------------|-------------|------------------|-------------|
| relaxation | EDH-mediated EDR | global EDR | EDH-mediated EDR | global EDR |
| No ASC | 99.1 ± 1.2 | 99.8 ± 2.8 | 98.5 ± 0.6 | 101.2 ± 1.9 |
| ASC 10 ⁻⁵ M | 84.1 ± 2.2* | 99.7 ± 3.8 | 90.5 ± 1.4* | 104.8 ± 3.9 |
| ASC 10 ⁻⁴ M | 34.5 ± 3.4* | 102.5 ± 2.7 | 51.7 ± 4.1* | 97.7 ± 4.2 |
| ASC 10 ⁻³ M | 7.5 ± 5.2* | 98.9 ± 4.1 | 10.5 ± 3.1* | 101.3 ± 3.6 |

*p < 0.05

EDR was induced by 10⁻⁵ M carbachol during contraction (first row in Table I) by PHE 10⁻⁵ M or PGF 10⁻⁵ M. EDR (second row in Table I) was examined either as global EDR or as its major part, EDH-mediated EDR. The influence of ascorbate (ASC, 10⁻⁵ M - 10⁻³ M) is shown in the last three rows (Table I). All ASC data are here normalised: EDR expressed as % of the respective EDR previously obtained in the same ring in the absence of ASC (mean ± SEM; n = 6 in all cases; *p < 0.01). The third row (ASC none) shows the reproducibility of that EDR in the same ring in the absence of ASC; values are used as the control for statistics. Our preliminary data showed carbachol 10⁻⁵ M induces complete global EDR and nearly maximal EDH-mediated EDR.

Within extensive research on EDR, studies also tested the relation between oxidative stress and NO-

mediated EDR, including the possible benefit from ASC. But here, we show that in isolated resistance arteries, ASC inhibits EDH-mediated EDR, the major EDR path in there. EDH-mediated EDR was inhibited by ASC in small mesenteric arteries (Table I). Still, the global EDR was not, probably because the reduced EDH-mediated EDR is compensated by the increased NO release by ASC shown by others [10]. Others showed ASC inhibits EDH-mediated EDR but used only perfused vascular beds [24, 25], then isolated larger arteries [14]. In the bovine ciliary bed and rat mesenteric bed, EDH-mediated EDR was blocked by physiological [ASC] [25]. They explored the same effect in isolated resistance mesenteric arteries, too [33]. Such studies focused on the relation between H₂O₂ and ASC as oxidative stress modulators, but then the EDHF role of H₂O₂ was not so well known.

Still, they did find that, in iliac artery rings, ASC can increase EDH-mediated EDR, by higher H_2O_2 [14], with its subsequent actions [27]. There they approached [14] ASC effect on the main path of EDH-mediated EDR, relying on hyperpolarisation propagating (via gap junctions) in both the endothelium and the VSM [13]. Typical resistance arteries, isolated, were tested in one study on ASC effects upon EDH-mediated EDR; in the model, we used here, they found no effect of ASC on EDH-mediated EDR [33]. Maybe because we used other contracting agents; see Table I vs. [33], while precontraction affects EDH-mediated EDR [36]. We used another way to compare test and control effects: separate tests for ASC 0.01, 0.1, and 100 mM; effects as % of the control in the same ring, limiting bias from other ASC effects. The flow was required for inhibition of EDH-mediated EDR by ASC [29, 33], but there was no flow in our tests and ASC still was able to inhibit EDH-mediated EDR (Table I).

We confirmed that ASC inhibits EDH-mediated EDR in rat mesentery [25], showing the effect is intrinsic to the vascular wall and resides distally, in accord with the increasing importance of EDH-mediated EDR along the arterial tree [36]. This ASC effect could be related to NOS facilitation by increased BH4 and/or a change in NO/EDHF balance (known ASC effects on NOS-NO-cGMP path). ASC can improve defective EDR by scavenging O_2^- , NO release from S-nitrosothiols, nitrite reduction to NO, and NOS activation [23]. ASC-induced endothelial recovery in atherosclerosis involves anti-oxidative NO protection, direct BH4 stabilisation, and better BH4-NOS operation; ASC endothelial saturation is required for BH4 protection and optimal NO synthesis [11]. VSM has large-conductance K_{Ca} (BK), an ascorbate target to protect against ischemic brain stroke [21]. The iK in endothelial projections are in a feedback loop with Ca_L in VSM [13, 28]; impaired NO-EDH cross-talk may be a therapeutic target in early endothelial dysfunction [1].

Conclusions

ASC reduces EDH-mediated EDR in resistance arteries. But ASC does not change the global EDR therein, probably due to the known effect of ASC on preserving and promoting NO-mediated EDR by opposing oxidative stress, but not only. EDH-mediated EDR should be considered in vascular studies on ASC. ASC addition to PSS for *in vitro* tests could be indicated to avoid discrepancies with *in vivo* studies, especially in studies regarding EDH-mediated EDR and oxidative stress.

Conflict of interest

The authors declare no conflict of interest.

References

1. Alaaeddine RA, Mroueh A, Gust S, Eid AH, Plane F, El-Yazbi AF, Impaired cross-talk between NO

- and hyperpolarisation in myoendothelial feedback: a novel therapeutic target in early endothelial dysfunction of metabolic disease. *Curr Opin Pharmacol.*, 2019; 45: 33-41.
2. Al-Khudairy L, Flowers N, Wheelhouse R, Ghannam O, Hartley L, Stranges S, Rees K, Vitamin C supplementation for the primary prevention of cardiovascular disease. *Cochrane Database Syst Rev.*, 2017; 3(3): CD011114.
3. Ashor AW, Lara J, Mathers JC, Siervo M, Effect of vitamin C on endothelial function in health and disease: a systematic review and meta-analysis of randomised controlled trials. *Atherosclerosis*, 2014; 235(1): 9-20.
4. Baker TA, Milstien S, Katusic ZS, Effect of vitamin C on the availability of tetrahydrobiopterin in human endothelial cells. *J Cardiovasc Pharmacol.*, 2001; 37: 333-338.
5. Behringer EJ, Hakim MA, Functional interaction among K_{Ca} and TRP channels for cardiovascular physiology: modern perspectives on ageing and chronic disease. *Int J Mol Sci.*, 2019; 20(6): E1380: 1-25.
6. Burnham MP, Bychkov R, Félétou M, Richards GR, Vanhoutte PM, Weston AH, Characterisation of an apamin-sensitive small-conductance Ca^{2+} -activated K^+ channel in porcine coronary artery endothelium: relevance to EDHF. *Br J Pharmacol.*, 2002; 135(5): 1133-1143.
7. Bychkov R, Burnham MP, Richards GR, Edwards G, Weston AH, Félétou M, Characterisation of a charybdotoxin-sensitive intermediate conductance Ca^{2+} -activated K^+ channel in porcine coronary endothelium: relevance to EDHF. *Br J Pharmacol.*, 2002; 137(8): 1346-1354.
8. Chen GC, Lu DB, Pang Z, Liu QF, Vitamin C intake, circulating vitamin C and risk of stroke: a meta-analysis of prospective studies. *J Am Heart Assoc.*, 2013; 2(6): e000329: 1-11.
9. Constantin M, Şerban DN, Pricop C, Huzum B, Şerban IL, Extracellular Mg^{2+} level affects the major mechanism of endothelium-dependent relaxation in resistance arteries. *Farmacia*, 2019; 67(5): 888-891.
10. de Saram K, McNeill KL, Khokher S, Ritter JM, Chowienzyk PJ, Divergent effects of vitamin C on relaxations of rabbit aortic rings to acetylcholine and NO-donors. *Br J Pharmacol.*, 2002; 135: 1044-1050.
11. d'Uscio LV, Milstien S, Richardson D, Smith L, Katusic ZS, Long-term vitamin C treatment increases vascular tetrahydrobiopterin levels and nitric oxide synthase activity. *Circ Res.*, 2003; 92: 88-95.
12. Félétou M, Endothelium-dependent hyperpolarisation and endothelial dysfunction. *J Cardiovasc Pharmacol.*, 2016; 67(5): 373-387.
13. Garland CJ, Bagher P, Powell C, Ye X, Lemmey HAL, Borysova L, Dora KA, Voltage-dependent Ca^{2+} entry into smooth muscle during contraction promotes endothelium-mediated feedback vasodilation in arterioles. *Sci Signal.*, 2017; 10(486): eaal3806: 1-15.
14. Garry A, Edwards DH, Fallis IF, Jenkins RL, Griffith TM, Ascorbic acid and tetrahydrobiopterin potentiate the EDHF phenomenon by generating hydrogen peroxide. *Cardiovasc Res.*, 2009; 84(2): 218-226.
15. Goto K, Ohtsubo T, Kitazono T, Endothelium-dependent hyperpolarisation (EDH) in hypertension: the role of

- endothelial ion channels. *Int J Mol Sci.*, 2018; 19(1): e315: 1-20.
16. Gradel AKJ, Salomonsson M, Sørensen CM, Holstein-Rathlou NH, Jensen LJ, Long-term diet-induced hypertension in rats is associated with reduced expression and function of small artery SK_{Ca}, IK_{Ca}, and Kir2.1 channels. *Clin Sci.*, 2018; 132(4): 461-474.
 17. He D, Pan Q, Chen Z, Sun C, Zhang P, Mao A, Zhu Y, Li H, Lu C, Xie M, Zhou Y, Shen D, Tang C, Yang Z, Jin J, Yao X, Nilius B, Ma X, Treatment of hypertension by increasing impaired endothelial TRPV4-KCa_{2.3} interaction. *EMBO Mol Med.*, 2017; 9(11): 1491-1503.
 18. Hearon CM Jr, Dinunno FA, Escape, lysis, and feedback: endothelial modulation of sympathetic vasoconstriction. *Curr Opin Pharmacol.*, 2019; 45: 81-86.
 19. Hogas S, Ardeleanu S, Segall L, Serban DN, Serban IL, Hogas M, Apetrii M, Onofriescu M, Sascau R, Covic A, Changes in arterial stiffness following dialysis in relation to overhydration and to endothelial function. *Int Urol Nephrol.*, 2012; 44(3): 897-905.
 20. Huzum B, Curpan AS, Puha B, Serban DN, Veliceasa B, Necoara RM, Alexa O, Serban IL, Connections between orthopedic conditions and oxidative stress: current perspective and the possible relevance of other factors, such as metabolic implications, antibiotic resistance, and COVID-19. *Medicina (Kaunas)*, 2022; 58(3): 439: 1-16.
 21. Li L, Li S, Hu C, Zhou L, Zhang Y, Wang M, Qi Z, BKCa channel is a molecular target of vitamin C to protect against ischemic brain stroke. *Mol Membr Biol.*, 2019; 35(1): 9-20.
 22. Maranduca MA, Tanase DM, Branisteanu DC, Serban DN, Branisteanu DE, Serban IL, Involvement of proinflammatory cytokines in angiotensin II-induced hypertension in rat. *Exp Ther Med.*, 2020; 20(4): 3541-3545.
 23. May JM, Harrison FE, Role of vitamin C in the function of the vascular endothelium. *Antioxid Redox Signal.*, 2013; 19(17): 2068-2083.
 24. McNeish AJ, Nelli S, Wilson WS, Dowell FJ, Martin W, Differential effects of ascorbate on endothelium-derived hyperpolarising factor (EDHF)-mediated vasodilatation in the bovine ciliary vascular bed and coronary artery. *Br J Pharmacol.*, 2003; 138(6): 1172-1180.
 25. McNeish AJ, Wilson WS, Martin W, Ascorbate blocks endothelium-derived hyperpolarising factor (EDHF)-mediated vasodilatation in the bovine ciliary vascular bed and rat mesentery. *Br J Pharmacol.*, 2002; 135(7): 1801-1809.
 26. Morelli MB, Gambardella J, Castellanos V, Trimarco V, Santulli G, Vitamin C and cardiovascular disease: an update. *Antioxidants (Basel)*, 2020; 9(12): 1227: 1-16.
 27. Muller-Delp JM, Ascorbic acid and tetrahydrobiopterin: looking beyond nitric oxide bioavailability. *Cardiovasc Res.*, 2009; 84(2): 178-179.
 28. Murphy TV, Sandow SL, Agonist-evoked endothelial Ca²⁺ signalling microdomains. *Curr Opin Pharmacol.*, 2019; 45: 8-15.
 29. Nelli S, Dowell FJ, Wilson WS, Stirrat A, Martin W, Requirement for flow in the blockade of endothelium-derived hyperpolarising factor (EDHF) by ascorbate in the bovine ciliary artery. *Br J Pharmacol.*, 2004; 142(7): 1081-1090.
 30. Niță D, Ionescu M, Mazilu L, Suceveanu AI, Munteanu A, Ionescu P, Tuță LA, Buicu F, Parepa IR, Statins and the risk for coronary in-stent restenosis in diabetic patients. *Farmacia*, 2021; 69(3): 576-584.
 31. Serban DN, Nilius B, Vanhoutte PM, The endothelial saga: the past, the present, the future. *Pflugers Arch.*, 2010; 459(6): 787-792.
 32. Serban DN, Serban IL, Advancement regarding the role of endothelium in arterial wall dysfunction. *Rev Med Chir Soc Med Nat Iasi.*, 2016; 120(2): 219-222.
 33. Stirrat A, Nelli S, Dowell FJ, Martin W, Flow-induced enhancement of constriction and blockade of endothelium-derived hyperpolarising factor (EDHF) by ascorbate in the rat mesentery. *Br J Pharmacol.*, 2008; 153(6): 1162-1168.
 34. Stoica BA, Bordeianu G, Stanescu R, Serban DN, Nechifor M, A new method for the quantification of superoxide dismutase mimics with an allopurinol-xanthine oxidase-lucigenin enhanced system. *J Biol Inorg Chem.*, 2011; 16(5): 753-761.
 35. Tilinca MC, Merlan I, Salcudean A, Tilea I, Nemes-Nagy E, Oxidative stress and cytokines' involvement in the occurrence and progression of diabetic complications in the COVID-19 pandemic context. *Farmacia*, 2021; 69 (4): 635-641.
 36. Tomioka H, Hattori Y, Fukao M, Sato A, Liu M, Sakuma I, Kitabatake A, Kanno M. Relaxation in different-sized rat blood vessels mediated by endothelium-derived hyperpolarising factor: importance of processes mediating precontractions. *J Vasc Res.*, 1999; 36: 311-320.