

## PROINSULIN AND ADIPONECTIN IN PATIENTS WITH NONALCOHOLIC FATTY LIVER DISEASE

ELENA CACEAUNE<sup>1\*</sup>, DANIELA LIXANDRU<sup>2</sup>, IONEL COPACI<sup>3</sup>,  
NICU CACEAUNE<sup>3</sup>, CONSTANTIN IONESCU-TÎRGOVIȘTE<sup>1</sup>,  
MOHAMAD ABDULWAHABE MOHAMAD<sup>2</sup>

<sup>1</sup>*National Institute of Diabetes, Nutrition and Metabolic Diseases "Prof. Dr. N. Paulescu", Bucharest, Romania*

<sup>2</sup>*University of Medicine and Pharmacy "Carol Davila" Bucharest, Romania*

<sup>3</sup>*Clinical Institute Fundeni, Bucharest, Romania*

\* *corresponding author: elena\_caceaune@yahoo.com*

### Abstract

Liver fat, more than visceral fat is associated with type 2 diabetes (T2D). Although the relationship of nonalcoholic fatty liver disease (NAFLD) was studied in the light of the metabolic syndrome and type 2 diabetes, the relationship of NAFLD with proinsulin and adiponectin levels reflecting the  $\beta$ -cells dysfunction and respectively adipocytes secretion was less studied. In 112 patients (41 men and 71 women) fasting plasma glucose, glycated hemoglobin (HbA1c), total cholesterol, HDL-cholesterol, triglycerides, fasting insulin, proinsulin and adiponectin were determined. Body fat percent and trunk fat content were measured using bioelectrical impedance analysis (TANITA BC-418). Weight, body mass index (BMI), waist circumference, adipose tissue percent, trunk adipose tissue percent, fasting insulin, homeostatic model assessment insulin resistance (HOMA-IR) and visceral fat ratio (VFR) were higher in patients with NAFLD than in patients without NAFLD, the highest value was found in patients with NAFLD and T2D. Proinsulin, proinsulin/insulin and proinsulin/adiponectin ratio were significantly higher in patients with NAFLD and T2D than all the other groups. Adiponectin was significantly lower in patients with than without NAFLD. Proinsulin/insulin ratio can be considered a marker of  $\beta$ -cells dysfunction only in patients with NAFLD and T2D and not in patients with NAFLD and without T2DM, emphasizing that proinsulin is not a marker of liver disease.

### Rezumat

Adipozitatea intrahepatică mai mult decât cea viscerală este asociată cu diabetul zaharat tip 2 (DZ tip 2). Deși s-a studiat relația steatozei hepatice nonalcoolice (SH) cu sindromul metabolic și diabetul zaharat tip 2, relația cu proinsulina și adiponectina, reflectând disfuncția  $\beta$ -celulară, și respectiv secreția adipocitară, a fost mai puțin studiată. La 112 pacienți (41 bărbați și 71 femei) s-au determinat: glicemia *a jeun*, HbA1c, colesterolul total, HDL-colesterolul, trigliceridele, insulinemia *a jeun*, proinsulina și adiponectina. Procentul de țesut adipos corporal și de la nivelul trunchiului s-au determinat prin analiza impedanței bioelectrice (TANITA BC-418). Greutatea, indicele de masă corporală (IMC), circumferința abdominală, procentul de țesut adipos corporal, cel de la nivelul trunchiului, insulinemia *a jeun*, *homeostatic model assessment insulin resistance* (HOMA-IR) și *visceral fat ratio* (VFR) au fost semnificativ mai mari la pacienții cu SH *versus* fără SH, valorile cele mai mari s-au obținut la pacienții cu SH și DZ tip 2.

Proinsulina, raportul proinsulină/insulină și proinsulină/adiponectină au fost semnificativ mai mari la pacienții cu SH și DZ tip 2 față de celelalte grupuri. Adiponectina a fost semnificativ mai mică la pacienții cu SH *versus* fără SH. Raportul proinsulină/insulină poate fi considerat un marker al disfuncției  $\beta$ -celulare numai la pacienții cu SH și DZ tip 2, nu și la pacienții cu SH fără DZ tip 2, subliniind că proinsulina nu este un marker al afectării hepatice.

**Keywords:** nonalcoholic fatty liver disease (NAFLD), type 2 diabetes, proinsulin, adiponectin.

### Introduction

Liver fat, more than visceral fat is associated with type 2 diabetes mellitus. Whether the liver fat precedes or follows the disturbance of blood glucose regulation, this is a question that has not been answered yet. Nonalcoholic fatty liver disease (NAFLD) emerged as an entity in the last decades, despite in 1920, in the subchapter “Obesity” from “*Traité de Physiologie Médicale*” [1] it was mentioned that in obesity “liver is voluminous and yellowish and have often its cells infiltrated with lipid granulations”. Already in 1912, in the third volume from the “*Traité de Médecine Laceraux-Paulesco*” [2], referring to the role of the liver in the regulation of blood glucose level, the “hepato-pancreas” was described as a binome organ: the internal secretion of the pancreas (not yet discovered at that time) influences the glycogenic function of the liver in maintaining posthepatic blood glucose levels in a normal range. In his previous studies Paulescu observed that after pancreatectomy in dogs, the capacity of liver in storing glycogen decreases. In this way he anticipated that the antidiabetic hormone secreted by the pancreas is essential for the glycogenic function of the liver.

Proinsulin concentration is low in normoglycemic subjects, having 10-20% of insulin activity regarding glucose lowering [3] and has an increased level in diabetic patients and in subjects with impaired glucose tolerance [4-12]. This is linked with a disturbance of the proinsulin processing inside  $\beta$ -cells [10-13]. Considering both impaired glucose tolerance (IGT) and type 2 diabetes (T2D), associated with overweight/obesity the increasing overload of  $\beta$ -cells induces a disproportionate proinsulin level relative to the insulin level. In the Insulin Resistance Atherosclerosis Study, Hanley et al. demonstrated that the decreasing of acute insulin response and the increasing of proinsulin concentration were significantly associated with the 5-year incidence of diabetes after being adjusted for initial glucose tolerance status and for insulin sensitivity. In a subgroup of subjects with

IGT the high level of proinsulin was associated with an increased risk of diabetes [14].

Adiponectin is the most abundant protein produced by adipose tissue with a higher level in females. Whereas the majority of proteins and free fatty acids secreted by adipose tissue increases with the increasing of visceral adiposity, the adiponectin level decreases [15]. Adiponectin increases the insulin sensitivity, free fatty acids oxidation, glucose uptake in skeletal muscle tissue, increases energy expenditure and reduces liver glucose production. Plasma levels vary from 2 to 10  $\mu\text{g/mL}$  being low in obese and type 2 diabetes patients, and in dyslipidemia. Also it is considered a good marker of cardiovascular risk. In the liver, muscular tissue and adipose tissue adiponectin increases insulin sensitivity [16, 17].

In a study of Kantartzis *et al*, using a state of art technics for determination of visceral fat and liver fat, it was found that the percentage of liver fat increases progressively from  $4.43\pm 0.32\%$  in normal glucose to  $5.85\pm 1.14\%$  in impaired fasting glucose (IFG), to  $7.97\pm 1.14\%$  in impaired glucose tolerance (IGT) and  $11.5\pm 1.44$  in IFG+IGT. They found that liver fat predicted the degree of glucose deterioration more accurately than total visceral fat. Also this study shows a decrease in adiponectin level from IFG to IGT then IFG and IGT, confirming previous data suggesting that this association depends on obesity [18, 19, 20]. Low levels of adiponectin in patients with liver steatosis are correlated with insulin sensitivity, liver fat content and also with inflammation and fibrosis. Moreover, decreased adiponectin level is considered a better predictor of hepatic steatosis and of the increasing level of ALT and GGT in obese persons without metabolic disorders [21, 22].

Although NAFLD was studied in connection with metabolic syndrome [23, 24, 25], cardiovascular diseases [26], and type 2 diabetes [27], the relationship of NAFLD with proinsulin and adiponectin levels reflecting the  $\beta$ -cells dysfunction and respectively adipocytes function was less studied.

Our study aimed to investigate the relation of proinsulin and adiponectin with nonalcoholic fatty liver disease according to the presence/absence of type 2 diabetes.

### Materials and Methods

Our study included 112 patients (41M/71F) with a mean age of  $55.22 \pm 8.95$  years old recruited from the I. Pavel Diabetes Center belonging to the National Institute of Diabetes, Nutrition and Metabolic Diseases "N. C. Paulescu", Bucharest, Romania, between March and December 2012.

NAFLD was diagnosed by ultrasonography. Exclusion criteria: positive serologic markers for viral hepatitis and alcohol consumption > 20g/day. All anthropometric measurements (weight, height, body mass index (BMI), waist circumference (WC) were done by the same physician on the day the blood sample was taken. Venous blood samples were drawn from all subjects after overnight fasting and routine blood tests including fasting plasma glucose (FPG), glycated hemoglobin (HbA1c), total cholesterol (TC), HDL-cholesterol, triglycerides (TG) and fasting insulin (FI) were analyzed. Proinsulin and adiponectin levels were determined by ELISA method using a commercial kit (DRG Instruments GmbH, Germany, EIA 1560 and EIA 4177). Total analytical variability, expressed as coefficient of variation (CV) was 4.3% and 6.8% respectively. BMI was calculated as the weight (kg) divided by square of the height (m). LDL-cholesterol was calculated with Friedewald's formula:  $TC - HDL-cholesterol - TG/5$  (for  $TG < 400$  mg/dL). We also calculated: Weight/Height (W/H) ratio, Proinsulin/Insulin ratio (P/I) and Proinsulin/Adiponectin ratio (P/A). Insulin sensitivity was calculated by HOMA-IR index, (*Homeostasis Model Assessment for Insulin Resistance*):  $[fasting\ plasma\ glucose\ (mmol/L) \times fasting\ insulin\ (\mu U/mL)]:22.5$  [28]. Individuals with HOMA-IR > 2.7 were considered as insulin resistant. Body fat percent and trunk fat content were measured using bioelectrical impedance analysis (TANITA BC-418).

The control group included 20 healthy subjects in which both NAFLD and T2D were excluded. The study protocol was approved by The Ethical Committee of "N.C. Paulescu" National Institute of Diabetes, Nutrition and Metabolic Disease, Bucharest, Romania. A written informed consent was obtained from each study participant.

*Statistical analysis:* Statistical Package for Social Sciences (SPSS 21.0) software (IBM). Kolmogorov-Smirnov test was used to analyze continuous data distribution, according to which appropriate tests were further used in analysis: ANOVA or Kruskal-Wallis test for differences between means of 4 independent groups. Spearman's correlation coefficients were calculated in order to test the association between variables. p-values < 0.05 were considered as statistically significant.

## Results and Discussion

The anthropometric and metabolic characteristics of the studied groups are shown in Table I. According to the presence or absence of NAFLD and T2D the cohort was divided into four groups: **group 1** – patients with T2D and without NAFLD (10.71 %); **group 2** – patients with

NAFLD and without T2D (17.86 %); **group 3** – patients with NAFLD and T2D (53.57 %) and the **control group** - subjects without NAFLD and T2D (17.86 %).

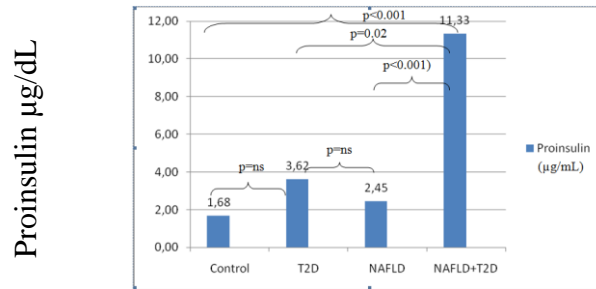
**Table I**  
Anthropometric and metabolic characteristics of the studied groups

	Control (n=20)	T2D (n=12)	NAFLD (n=20)	NAFLD+T2D (n=60)	p-value
Age (years)	42.80±10.87	59.42±0.63	49.70±8.42	56.73±7.59	<0.001*
Weight (kg)	66.06±13.14	66.93±11.00	83.11±13.96	88.65±15.34	<0.001*
BMI (kg/m <sup>2</sup> )	24.33±3.95	24.84±1.91	29.89±4.57	32.72±4.89	<0.001*
WC (cm)	83.15±11.69	93.75±6.68	101.55±10.96	108.73±9.97	<0.001*
FPG (mg/dL)	91.51±8.31	137.19±48.23	102.13±12.20	146.71±38.65	<0.001*
HbA1c (%)	5.28±0.42	6.54±0.56	5.62±0.31	7.09±1.36	<0.001**
TC (mg/dL)	204.54±52.52	208.24±43.44	235.44±41.74	206.38±45.26	0.099*
HDL-c (mg/dL)	56.81±10.99	55.30±16.47	49.39±13.40	46.38±12.49	0.008*
LDL-c (mg/dL)	129.87±52.54	125.65±39.60	151.80±47.82	126.22±41.84	<0.001*
Triglycerides (mg/dL)	89.57±35.97	150.45±81.53	189.33±133.83	183.47±105.89	<0.001*
Fasting insulin (μU/mL)	7.99±3.31	8.61±2.13	11.68±6.86	16.62±9.91	<0.001**
HOMA-IR index	1.83±0.85	3.10±1.22	3.05±2.08	5.99±3.64	<0.001**
Proinsulin (μU/mL)	1.68±1.40	3.62±2.58	2.45±1.96	11.33±13.99	<0.001**
PI/I ratio	0.20±0.19	0.39±0.24	0.22±0.23	0.69±0.62	<0.001**
Adiponectin (ng/mL)	12.31±6.92	13.90±7.63	9.75±8.84	7.33±4.86	0.005*
PI/A ratio	0.26±0.42	0.38±0.41	0.48±0.45	5.58±12.90	<0.001**
% Adipose tissue	27.50±9.34	26.37±7.77	35.28±8.22	37.97±8.41	<0.001*
% Trunk adipose tissue	25.40±9.87	23.67±7.90	34.59±7.31	35.67±7.18	<0.001*
VFR	5.41±2.85	8.62±2.92	10.66±4.25	12.55±3.17	<0.001*

\*ANOVA; \*\*Kruskal-Wallis; BMI: body mass index; WC: waist circumference; TC: total cholesterol; FPG: fasting plasma glucose; FI: fasting insulin. VFR: visceral fat ratio; PI/I: Proinsulin/Insulin; PI/A: Proinsulin/Adiponectin

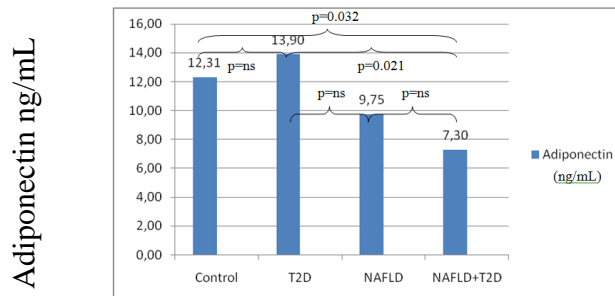
Weight, BMI, waist circumference, adipose tissue percent, trunk adipose tissue percent, HOMA-IR and VFR were higher in patients with NAFLD than in patients without NAFLD, the highest value was found in patients with NAFLD and T2D. Moreover, in patients with NAFLD, fasting insulin was significantly higher than in patients without NAFLD, while HDL-cholesterol was significantly lower in patients with NAFLD comparing with control group.

As shown in Figure 1, proinsulin was significantly higher in patients with NAFLD and T2D (11.33  $\mu\text{g/dL}$ ) than all other groups included in our study (1.68  $\mu\text{g/dL}$  – control group; 3.62  $\mu\text{g/dL}$  – group 1 and 2.45  $\mu\text{g/dL}$  – group 2 respectively). Adiponectin was significantly lower in patients with (7.3 ng/mL) than without NAFLD (12.31 ng/mL; 13.90 ng/mL), (Figure 2). For proinsulin/insulin ratio (Figure 3) and proinsulin/adiponectin ratio (Figure 4) we found higher level in patients which associated NAFLD and T2D.



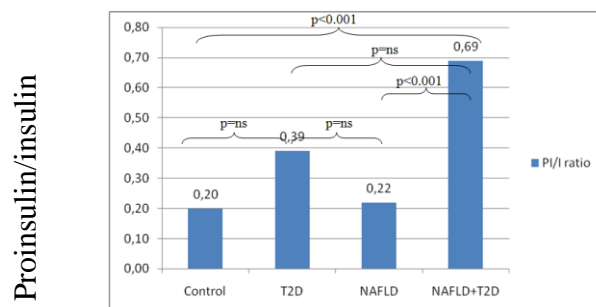
**Figure 1.**

The measured values of proinsulin in the studied groups



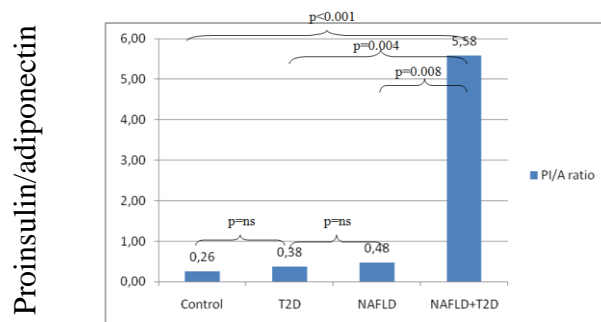
**Figure 2.**

The measured values of adiponectin in the studied groups



**Figure 3.**

The proinsulin/insulin ratio in the studied groups



**Figure 4.**

The proinsulin/adiponectin ratio in the studied groups

In patients with NAFLD and T2D **proinsulin** was correlated with weight, BMI, waist circumference, VFR, fasting plasma glucose, HbA1c, HOMA-IR and triglycerides; **adiponectin** was positively correlated with HDL-cholesterol and negatively with waist circumference and fasting insulin; **proinsulin to insulin ratio** was positively correlated with VFR, FPG, HbA1c; **proinsulin to adiponectin** ratio was positively correlated with weight BMI, waist circumference, VFR, fasting insulin, HOMA-IR and triglycerides (Tabel II). After adjusting for BMI and waist circumference the correlations between P/A ratio with FI and HOMA-IR remained significantly statistic.

**Table II**

The correlations coefficients between the studied parameters in patients with NAFLD and T2D

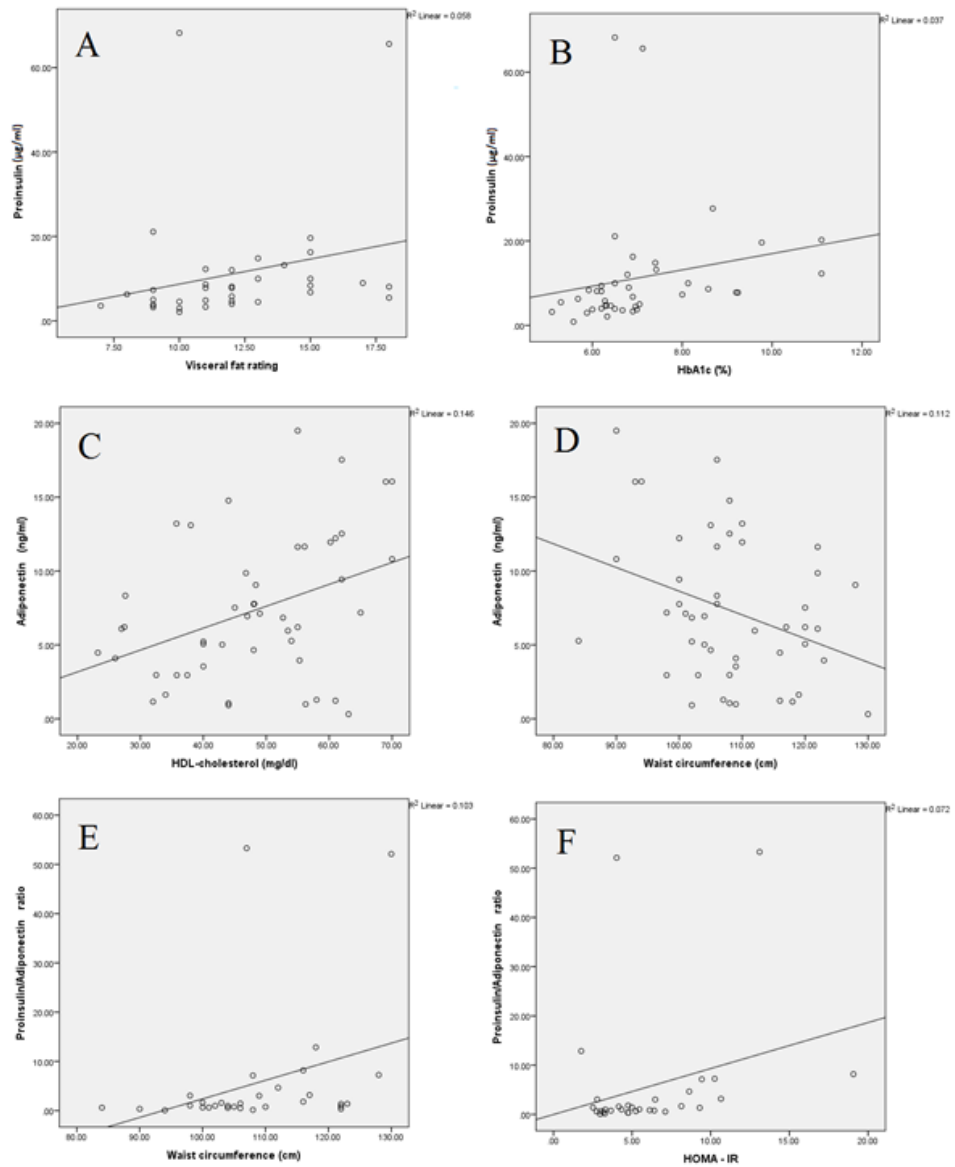
	Proinsulin	Adiponectin	Proinsulin/ Insulin	Proinsulin/ Adiponectin
<b>Weight</b>	0.397*	-0.273	0.233	0.379*
<b>BMI</b>	0.348*	-0.249	0,090	0.463**
<b>WC</b>	0.359*	-0.335	0,089	0.452*
<b>VFR</b>	0.471**	-0.211	0.451**	0.394*
<b>FPG</b>	0.346*	0.016	0.387*	0.134
<b>HbA1c</b>	0.514**	0.036	0.480**	0.347
<b>FI</b>	0.295	-0.343*	-0.241	0.387*
<b>HOMA-IR</b>	0.412**	-0.310*	-0.072	0.418*
<b>Triglycerides</b>	0.522**	-0.185	0.287	0.382*
<b>HDL-c</b>	-0.242	0.381**	-0.151	-0.143

\*\* Correlation is significant at the 0.01 level (2-tailed).

\* Correlation is significant at the 0.05 level (2-tailed).

In patients with NAFLD and without T2D we found a positive correlation for **proinsulin** with FPG ( $\rho=0.496$ ,  $p<0.05$ ) and triglycerides

( $\rho=0.484$ ,  $p<0.05$ ) and for **adiponectin** with HDL-cholesterol ( $r=0.587$ ,  $p<0.01$ ).



**Figure 5.**

The correlations between the studied parameters: proinsulin and visceral fat rating (A), proinsulin and HbA<sub>1c</sub> (B), adiponectin and HDL-cholesterol (C), adiponectin and waist circumference (D), proinsulin/adiponectin ratio and waist circumference (E), proinsulin/adiponectin ratio and HOMA-IR (F)



Liver is an important organ involved in the regulation of energy metabolism especially in the energy overflow induced by a high caloric/fat intake and the decrease of the physical activity. Placed between the pancreas and the adipose tissue, the liver is influenced on the one site by the insulin secretion (the portal vein contains the highest level of insulin) and on the other site by the high flow of the fatty acids released from lipolysis in the adipose tissue. Also, the liver receives not only adiponectin (protective hormone) but the proinflammatory cytokines as well, produced either by “stressed and aggressive” adipocytes [29, 30] or by the macrophages (TNF- $\alpha$ , IL-6) which invade the adipose tissue when the diameter of adipocytes overpass a limit (more than 110 $\mu$ m diameter) [31, 32, 33].

The most common conditions associated with NAFLD are obesity, T2D and dyslipidemia, liver fat being correlated with waist circumference and metabolic syndrome features [34]. NAFLD precedes and predicts T2D and cardiovascular diseases, independent of obesity [35]. Fasting insulin and HOMA-IR index in patients which associated T2D and fatty liver, are better correlated with total body adipose tissue than with trunk adiposity [36].

In our study the association of NAFLD and T2D was characterized by high values of BMI, WC, % body adipose tissue, % trunk adipose, VFR, insulin and HOMA-IR values.

Increased proinsulin and proinsulin/insulin ratio (marker of disproportionate release of proinsulin from  $\beta$ -cells), are correlated with insulin response being markers of  $\beta$ -cells secretory dysfunction [37, 38]. In patients with NAFLD and T2D proinsulin level was increased and correlated with anthropometric measures (weight, BMI, WC, VFR) and also with metabolic parameters (FPG, HbA<sub>1c</sub>, TG and HOMA-IR). Although NAFLD is considered to be a slowly progressive disease, the progression to various degrees of fibrosis is unexpectedly high. In a study of 103 patients who underwent serial liver biopsies (medium-interval of 3.2 years) fibrosis progressed in 37%, remained stable in 34% and regressed 29% in of the cases. The higher rate of progression has been noted in T2D patients or in patients with an initial even low degree of fibrosis [39]. In our study patients with NAFLD and without T2D had higher values of fasting insulinemia and HOMA-IR index compared with the control group, while proinsulin and proinsulin/insulin ratio were not found to be significantly changed. These data might be related to a decrease in the clearance of insulin related to the fat deposit within this organ. In this group, proinsulin was significantly correlated with FPG and TG. These data are consistent with those in a study performed on 64 Japanese type 2 diabetic subjects that have found a

significant positive correlation of proinsulin with BMI, hypertension, glucose, HbA1c, insulin and HOMA-IR. Proinsulin/Insulin ratio showed a significant positive correlation with glucose, HbA1c and with advanced glycation endproducts, as a consequences of glucotoxicity and causative for beta cells dysfunction and a significant negative correlation with insulin and HOMA-IR [13].

Mykkanen *et al* [40] showed that fasting insulin and intact proinsulin were inversely correlated and proinsulin-to-insulin ratio positively correlated with insulin sensitivity. Proinsulin/insulin ratio was inversely correlated with acute insulin response. The associations were independent of age, sex, BMI. The relation of acute insulin response with proinsulin-to-insulin ratio was independent of insulin sensitivity. Normoglycemic subjects with a low acute insulin response had a high level of proinsulin-to-insulin ratio compared with the patients with high acute insulin response and low proinsulin-to-insulin ratio. This can be explained by intensive processing of proinsulin in subjects with normal glucose. In normoglycemic subjects with insulin resistance proinsulin related to insulin level are not increased [40].

In a cross-sectional study on subjects without diabetes it was observed that proinsulin levels and proinsulin/insulin ratio increased with age both in men and women after adjusting for glucose and insulin sensitivity suggesting that  $\beta$ -cells' function decreases with age [41]. In our study, proinsulin and proinsulin to insulin ratio are significantly higher in patients with NAFLD only in association with T2DM, before and after adjusting for age, suggesting that the  $\beta$ -cells secretory dysfunction is not present in patients with NAFLD and without T2D.

It is well known that between plasma levels of adiponectin and the risk of obesity, diabetes and cardiovascular diseases exists a negative correlation, adiponectin concentration being negatively correlated with adipose tissue mass [16, 17].

Adiponectin levels can predict the presence of metabolic syndrome. The level of adiponectin was positively correlated with age, female gender, HDL-cholesterol and negatively with WC, BMI, insulin resistance, triglycerides and aminotransferase levels. In patients with NAFLD the plasma level of adiponectin is low and is correlated with increased age, female gender and triglycerides and inversely correlated with the percentage of hepatic fat content [42]. Bugianesi and Pagano showed that the adiponectin level in patients with NAFLD is 20-60% lower than in normal subjects [42, 43]. Moreover, Speliotes *et al* [44] demonstrated that NAFLD is associated with metabolic syndrome and with a low level of adiponectin independent of BMI, waist to hip ratio and visceral obesity. Our data

support those observations: adiponectin had the lowest value in NAFLD independent of the presence of T2D.

Proinsulin/adiponectin ratio as a marker of insulin resistance [45] was correlated in patients with NAFLD and T2D with anthropometric measures: weight, BMI, WC, VFR and also with insulin level, HOMA-IR index and triglycerides.

### Conclusions

The relationship between T2D and NAFLD could be bilateral, metabolic disorders may cause NAFLD and NAFLD may enhance metabolic disorders of diabetes.

Proinsulin had the highest values in patients with NAFLD and T2D. Proinsulin/insulin ratio can be considered a marker of  $\beta$ -cells dysfunction in patients with NAFLD and T2D but not in patients with NAFLD and without T2D, emphasizing that proinsulin is a marker of  $\beta$ -cells dysfunction and not a marker of liver disease.

The studied subjects presented decreased levels of adiponectin in NAFLD patients independent of the presence of T2D. Proinsulin/adiponectin ratio had the highest values in patients with NAFLD and T2D which may have a disturbance of  $\beta$ -cells function related to the excess of adipose tissue.

The early diagnosis and appropriate treatment of diabetes might represent the best way to prevent the progression of NAFLD.

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### References

1. Paulescu N. C.: *Traite de Physiologie Medicale*, 3 vol., 2210 pag., Bucharest, 1919-1921.
2. Lancereaux E. Paulescu N., *Traité de Médecine Laceraux-Paulesco*, 1912; vol.III, Paris.
3. Pfützner A, Forst T. Elevated intact proinsulin levels are indicative of Beta-cell dysfunction, insulin resistance, and cardiovascular risk: impact of the antidiabetic agent pioglitazone. *Journal of Diabetes Science and Technology*. 2011 May 1; 5(3): 784-793.
4. Reaven GM, Chen Y-DI, Hollenbeck CB, Sheu WHH, Ostrega D, Polonsky KS: Plasma insulin, C-peptide, and proinsulin concentrations in obese and

- nonobese individuals with varying degrees of glucose tolerance. *J Clin Endocrinol Metab* 1993; 76: 44-48.
5. Birkeland KI, Torjesen PA, Erikson J, Vaaler S, Groop L: Hyperproinsulinemia of type II diabetes is not present before the development of hyperglycemia. *Diabetes Care* 1994; 17: 1307-1310.
  6. Davies MJ, Rayman G, Gray IP, Day JL, Hales CN: Insulin deficiency and increased plasma concentration of intact and 32/33 split proinsulin in subjects with impaired glucose tolerance. *Diabet Med* 1993; 10: 313-320.
  7. Haffner SM, Stern MP, Miettinen H, Gingerich R, Bowsher RR: Higher proinsulin and specific insulin are both associated with a parental history of diabetes in nondiabetic Mexican-American subjects. *Diabetes* 1995; 44: 1156-1160.
  8. Popescu A.L., Virgolici B., Păcurar D., Timnea O., Ranetti A.E., Orășeanu D., Zăgrean L., Beneficial effects of omega-3 fatty acids in nonalcoholic fatty liver disease, in childhood obesity, *Farmacia*, 2013; 61(3): 598-608.
  9. Mykkanen L, Haffner SM, Kuusisto J, Pyorala K, Hales CN, Laakso M: Serum proinsulin levels are disproportionately increased in elderly prediabetic subjects. *Diabetologia* 1995; 38: 1176-1182.
  10. Nypels G, Popp-Snyders C, Kostense PJ, Bouter LM, Heine RJ: Fasting proinsulin and 2-h post-load glucose levels predict the conversion to NIDDM in subjects with impaired glucose tolerance: the Hoorn Study. *Diabetologia* 1996; 170: 113-118.
  11. Beer SF, O'Rahilly S, Spivey RS, Hales CN, Turner RC: Plasma proinsulin in first-degree relatives of type 2 diabetic subjects. *Diabetes Res* 1990; 14: 51-54.
  12. Hales CN, Byrne CD, Petry CJ, Wareham NJ: Measurement of insulin and proinsulin. *Diabetes Rev* 1996; 4: 320-335.
  13. Saisho Y, Maruyama T, Hirose H, Saruta T. Relationship between proinsulin-to-insulin ratio and advanced glycation endproducts in Japanese type 2 diabetic subjects. *Diabetes Res Clin Pract.* 2007 Nov; 78(2): 182-188. Epub 2007 Apr 30.
  14. Hanley AJ, D'Agostino R Jr, Wagenknecht LE, Saad MF, Savage PJ, Bergman R, Haffner SM. Increased proinsulin levels and decreased acute insulin response independently predict the incidence of type 2 diabetes in the insulin resistance atherosclerosis study. *Diabetes*. 2002; Apr; 51(4): 1263-1270.
  15. Aguilar-Salinas CA, García EG, Robles L, Riaño D, Ruiz-Gomez DG, García-Ulloa AC, Melgarejo MA, Zamora M, Guillen-Pineda LE, Mehta R, et al. High adiponectin concentrations are associated with the metabolically healthy obese phenotype. *J Clin Endocrinol Metab*; 2008; 93: 4075-4079.
  16. Galic Sandra, Oakhill Jon S., Steinberg Gregory R. Adipose tissue as an endocrine organ. *Molecular and Cellular Endocrinology* 2010; 316: 129-139.
  17. Fonseca-Alaniz Miriam Helena, Takada Julie, Cardoso Alonso-Vale Maria Isabel, Bessa Lima Fabio. Adipose tissue as an endocrine organ: from theory to practice. *Journal de Pediatria*, 2007; 83(5 Suppl): S192-203.
  18. Kantartzis K., Fritsche A, Tschritter O et al. The association between plasma adiponectin and insulin sensitivity in humans depends on obesity. *Obes Res* 2005; 13: 1683-1691.
  19. Kantartzis K., Peter A., Machicao F., Machann J., Wagner S., Konigsrainer I., Konigsrainer A., Schick F., Haring H-U., Stefan N.: Dissociation between fatty liver and insulin resistance in humans carrying a variant of the patatin-like phospholipase 3 gene. *Diabetes* 2009; 58: 2616-2623.

20. Szczepaniak LS, Nurenberg, Leonard D et al. Magnetic resonance spectroscopy to measure hepatic triglycerides content: prevalence of hepatic steatosis in the general population. *Am J Physiol Endocrinol Metab* 2005; 288: E462-E468.
21. Targher G, Bertolini L, Scala L, Poli F, Zenari L, Falezza G. Decreased plasma adiponectin concentrations are closely associated with nonalcoholic hepatic steatosis in obese individuals. *Clin Endocrinol (Oxf)* 2004; 61: 700–703.
22. Hui JM, Hodge A, Farrell GC, Kench JG, Kriketos A, George J. Beyond insulin resistance in NASH: TNF-alpha or adiponectin? *Hepatology*. 2004; 40: 46–54.
23. Kotronen A, Westerbacka J, Bergholm R, Pietilainen KH, Yki-Jarvinen H. Liver fat in the metabolic syndrom. *J Clin Endocrinol Metab* 2007; 92: 3490-3497.
24. Despres J. P., Lemieux I.: Abdominal obesity and metabolic syndrome. *Nature* 2006; 444: 881-887.
25. Caceaune E., Licarioiu D., Bradescu O., Caceaune N., Ionescu-Tîrgoviste C. Metabolic disorders and cardiovascular risk enhanced by nonalcoholic fatty liver disease in patients with metabolic syndrome. *Romanian Journal of Diabetes, Nutrition and Metabolic Diseases*, volume 19 (2012) / No. 2 / April-June.
26. Targher G., Marra F., Marchesini G. Increased risk of cardiovascular disease in non-alcoholic fatty liver disease: causal effect or epiphenomena? *Diabetologia* 2008; 51: 1947-1953.
27. Roden M. Mechanisms of disease: hepatic steatosis in type 2 diabetes-pathogenesis and clinical relevance. *Nat. Clin. Pract Endocrinol Metab* 2006; 2: 335-348.
28. Matthews DR., Hosker JP., Rudenski AS., Naylor BA., Treacher DF., Turner RC – Homeostasis model assessment: insulin resistance and  $\beta$ -cell function from fasting plasma glucose and insulin concentrations in man. *Diabetologia* 1985; 28: 412-419.
29. Ionescu-Tirgoviste C Insulin Resistance - What is Myth and What is Reality? *Acta Endo (Buc)* 2011; 7 (1): 123-146, doi:10.4183/aeb.2011.123.
30. Ionescu-Tîrgoviște C., C. Guja, The relationship between proinsulin level and body mass index in various diabetic phenotypes, including obesity and the metabolic syndrome. *Diabetes Vasc Dis Res* 2007; 4 (Suppl.1): S 109.
31. Spalding K.L.: Retrospective birth dating of cells in humans. *Cell* 2005; 122: 133-143.
32. Spalding K.L., Arner E., Westermark P.O. et al.: Dynamics of fat cell turnover in humans. *Nature* 2008; 453: 783-787.
33. Gligor R., Pușchiță M., Zdremțan D., Crîsnic I., Ionescu-Tîrgoviște C., Relationship between adiponectin and some metabolic parameters in obese and diabetic patients, *Farmacia*, 2012; 60(2): 293-306.
34. Caceaune E. The relation between nonalcoholic fatty liver disease and the metabolic syndrome. *Proceedings of the Romanian Academy*, Series B, 2012; vol. 2: 143-150.
35. Kotronen A., Yki-Jarvinen H. - Fatty Liver –A Novel Component of the Metabolic Syndrome, *Arterioscler Thromb Vasc Biol*. 2008; 28: 27-38.
36. Caceaune E., Mihai A., Licarioiu D., Copaci I., Caceaune N., Ionescu-Tirgoviste C. Nonalcoholic fatty liver disease: a marker of adipose tissue distribution, *The 3<sup>rd</sup> International Symposium on Adipobiology and Adipopharmacology (ISSA)*, 25-27 Oct. 2012, Burgas.

37. Shiraishi I, Iwamoto Y, Kuzuya T, Matsuda A, Kumakura S: Hyperinsulinaemia in obesity is not accompanied by an increase in serum proinsulin/insulin ratio in groups of human subjects with and without glucose intolerance. *Diabetologia* 1991; 34: 737-741.
38. Pradhan AD, Manson JE, Meigs JB, Rifai N, Buring JE, Liu S, Ridker PM. Insulin, proinsulin, proinsulin:insulin ratio, and the risk of developing type 2 diabetes mellitus in women. *The American Journal of Medicine* 2003 Apr 15; 114(6): 438-444.
39. Adams LA, Angulo P. Recent concepts in non-alcoholic fatty liver disease. *Diabet Med.* 2005 Sep; 22(9): 1129-1133.
40. Mykkänen L, Haffner SM, Hales CN, Ronnema T, Laakso M. The relation of proinsulin, insulin, and proinsulin-to-insulin ratio to insulin sensitivity and acute insulin response in normoglycemic subjects. *Diabetes.* 1997; 46(12): 1990-1995.
41. Bente Bryhni, Egil Arnesen, Trond G Jenssen. Associations of age with serum insulin, proinsulin and the proinsulin-to-insulin ratio: a cross-sectional study. *BMC Endocrine Disorders* 2010; 10:21.
42. Bugianesi E, Pagotto U, Manini R, Vanni E, Gastaldelli A, de Iasio R, Gentilcore E, Natale S, Cassader M, Rizzetto M, Pasquali R, Marchesini G. Plasma adiponectin in nonalcoholic fatty liver is related to hepatic insulin resistance and hepatic fat content, not to liver disease severity. *J Clin Endocrinol Metab.* 2005 Jun; 90(6): 3498-3504. Epub 2005 Mar 29.
43. Pagano C, Soardo G, Esposito W, et al.: Plasma adiponectin is decreased in nonalcoholic fatty liver disease. *Eur J Endocrinol.* 2005; 152(1): 113-118.
44. Speliotes EK, Massaro JM, Hoffmann U, Vasan RS, Meigs JB, Sahani DV, Hirschhorn JN, O'Donnell CJ, Fox CS. Fatty liver is associated with dyslipidemia and dysglycemia independent of visceral fat: the Framingham Heart Study. *Hepatology* 2010; 51: 1979-1987.
45. Langenfeld MR, Forst T, Standl E, Strotmann HJ, Lübber G, Pahler S, Kann P, Pfützner A; IRIS II study: sensitivity and specificity of intact proinsulin, adiponectin, and the proinsulin/adiponectin ratio as markers for insulin resistance. *Diabetes Technol Ther.* 2004; 6(6): 836-843.

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