POSSIBLE MOLECULAR MECHANISMS AND PATHWAYS INVOLVED IN BH3 MIMETIC ACTIVITY OF ALPHA-LIPOIC ACID ON HUMAN COLON CANCER CELL LINE

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Abstract

Alpha-lipoic acid (ALA), a naturally-occurring antioxidant, inhibits proliferation and induces apoptosis in various cancer cell lines without effects on normal non-transformed cells. The aim of this study was to examine the effects of alpha-lipoic acid (ALA), alone and combined with 5-fluorouracil (5-FU), on Bcl-2/Bax expression in human colon cancer Caco-2 cell line as well as to investigate possible molecular mechanisms and pathways involved in ALA-mediated effects. In the present study ALA and 5-FU showed a tendency to decrease Bcl-2 and increase Bax expression. ALA exerted higher inhibitory effects on Bcl-2 expression, while the significant increase of Bax expression was shown after the treatment with the combination of ALA and 5-FU. The binding modes of ALA and 5-FU with both targets were shown to be closely similar, and some interactions the same like those of known BH3 mimetics. Thus, ALA may be considered as potential BH3 mimic. Additionally, with in silico calculated physico-chemical properties taken into account, it was confirmed that ALA may easily be delivered to its intracellular and membrane targets.

Keywords: alpha-lipoic acid, BH3 mimetic, colon cancer, molecular mechanisms

Introduction

Alpha-lipoic acid (ALA), a small dithiol molecule derived from octanoic acid, may act as a powerful micronutrient with diverse pharmacological and anti-oxidant properties [1]. This naturally-occurring anti-oxidant, known as a vitamin, co-factor of some mitochondrial enzymes (pyruvate dehydrogenase, α-ketoglutarate dehydrogenase, glycine decarboxylase), potent free radical scavenger and drug candidate, may be de novo synthesized in small amounts by animals and humans [2, 3]. It can also be absorbed from dietary sources such as red meat, potatoes and may be present in wheat and, to a lesser degree, in fruits and vegetables [4, 5]. Namely, high ALA contents (0.55 to 2.36 ppm) are present in food of animal origin (mainly in liver and muscles), whereas plants contain very little (0.09 ppm) or no detectable ALA [6]. It has been used like a dietary supplement in the prevention or treatment of stroke, diabetes, neuro-degenerative and hepatic disorders [3]. Moreover, it has been reported that ALA inhibits proliferation and induces apoptosis in various cancer cell lines including human lung epithelial cancer NCI-H460 cells [7], human cervical carcinoma HeLa S3 cells [8].
ovarian carcinoma cell lines IGROV1 and IGROV1-R10 [9], SMMC-7721 human hepatoma cell line [10], MDA-MB-231 breast cancer cells [11], T24 human bladder cancer cell line [12] and human promyelocytic HL-60 cells [3], without effects on normal non-transformed cells [7]. The cytotoxicity of ALA at millimolar concentrations was shown on SW620 human colon carcinoma cells as well as the enhancement of this effect in a synergistic manner in the combination with ascorbate [13]. It has been determined that ALA is able to effectively induce apoptosis in human colon cancer HT-29 cells via increased ROS (reactive oxygen species) production in mitochondria [14]. ALA has been reported to induce cell death in HT-29 and Caco-2 cell lines mediated by the activation of caspase-9, -3 and -7, and potentiate the cytotoxicity of 5-fluorouracil (5-FU) in these cells [15]. ALA may be considered as potential novel drug candidate for cancer therapy, but mechanisms of its chemopreventive effects need to be better clarified.

Continuing research on chemopreventive potential of ALA, after testing its effects alone or in combination with 5-FU or cisplatin on the proliferation of colon cancer cell lines and proved anti-proliferative activity [8], the aim of this study was to assess the effects of ALA, alone and combined with 5-FU, on Bel-2/Bax expression in human colon cancer Caco-2 cell line as well as to investigate possible molecular mechanisms and pathways involved in ALA-mediated effects. Molecular docking studies were performed to gain an insight into the binding modes of possible interactions of ALA and 5-FU with Bcl-2 and Bax proteins as checkpoints of intrinsic apoptotic pathway.

Additionally, an in silico study using Molinspiration tool [16] was performed in order to gain a better insight into the physico-chemical properties of ALA.

Materials and Methods

Chemicals

Alpha-lipoic acid (ALA) and 5-fluorouracil (5-FU) were purchased for experiments as follows: ALA (Berliton ED 300, Berlin-Chemie, Germany, 300 mg/12 mL) and 5-FU (Fluourouracil Teva, Pharmachemie BV - Netherlands, 50 mg/mL). DMEM (Dulbecco's Modified Eagle Medium), FBS (Foetal Bovine Serum), antibiotic/ antimycotic solution, L-glutamine and Trypsin-EDTA solution were purchased from PAA Laboratories (PAA Laboratories, Austria). Trypan blue for cell staining was purchased from Invitrogen. Primary anti-Bcl-2 and anti-Bax antibodies and secondary antibodies were purchased from Santa Cruz Biotechnology (Santa Cruz, CA, USA). Examined cytostatic drug 5-FU is used in the treatment of colon carcinoma according to the protocol [17].

Cell line

In this study we used Caco-2 cell line (human colon cancer cells) which was obtained from ATCC. Cells were cultured in DMEM supplemented with 10% FBS, 2 mM L-glutamine and antibiotic/antimycotic solution at 37°C in an atmosphere with 5% CO₂ and saturated humidity. Replacement of the culture medium was performed every 2 to 3 days.

Treatment of cells

Confluent culture of Caco-2 cells was harvested using Trypsin-EDTA solution, washed in buffer solution and the total number of cells was determined by Trypan blue dye exclusion test. Cells were seeded in 96-well plates (Greiner Bio-One, Germany) at a density of 3 x 10⁴ cells per well and cultured for 24 h under standard cell culture conditions. After that, the examined compounds, alone or in combination, were added to the cells. ALA and 5-FU were diluted in DMEM and three concentrations of each of these compounds were tested (group 1 – the lowest concentration, group 2 – middle concentration and group 3 – the highest concentration). Final concentrations of the assayed compounds were 10, 100 and 1000 μM. Combining of ALA and 5-FU was performed using the same concentrations as the following: group 1 with group 1, group 2 with group 2, and group 3 with group 3 in the ratio 1:1, so that the effective concentrations of compounds in combinations were twice less than the concentrations of compounds that were applied alone. As control we used cells that were incubated only with completed cell culture medium, DMEM, without the assayed compounds. Cells were incubated with ALA and 5-FU for the next 48 h. After that, the level of Bcl-2 and Bax proteins was measured.

Measurement of Bcl-2 and Bax protein levels

For determining the levels of Bcl-2 and Bax proteins, the cells were treated as it was described in the section “Treatment of cells”. After 48 h of incubation with the assayed compounds cells were further processed according to the protocol by Kocic et al. [18]. Briefly, the cells were washed with phosphate-buffered saline (PBS), fixed by using 70% methanol and permeabilized with 0.1% Triton in PBS. The cells were incubated with the primary anti-Bax and anti-Bcl-2 antibodies, washed three times and incubated with the FITC-conjugated secondary antibodies. The mean fluorescence intensity (MFI; logarithmic scale) was determined and analysed on a Victor™ multipe plate reader (Perkin Elmer-Wallace, Wellesley, MA). The presented results were obtained following the subtraction of blank values obtained by the treatment with the secondary antibodies only.

Ligand and receptor preparation and validation of docking protocol

The molecular docking study was performed using MOE 2014.0901 to understand the ligand protein interactions in detail. The X-ray crystallographic structures of Bcl-2 protein complexed with inhibitor (PDB code: 4IEH) and Bax protein complexed with
activator (PDB code: 2K7W) were obtained from the Protein Data Bank [19, 20].

**Statistical analysis**

The data were analysed by the commercially available statistics software package (SPSS for Windows®, v. 17.0, Chicago, USA) using the Students’ t-test and the ANOVA test. The results are presented as percentage of control. The statistical significance was set to p < 0.05.

**Results and Discussion**

Figure 1 shows the levels of Bcl-2 and Bax quantitative expression in Caco-2 cell culture treated with different concentrations of ALA and 5-FU under the given experimental conditions. Both tested compounds showed a tendency to decrease Bcl-2 and increase Bax expression levels, compared with control samples. It was shown that ALA exerts a significant inhibitory effect on Bcl-2 expression at the concentration of 1000 µM, while the effect of 5-FU at the same concentration was lower. The significant increase of Bax expression was shown after the treatment with 5-FU alone and its combination with ALA, especially at the highest tested concentrations of both compounds. Literature data showed that 5-FU-induced apoptosis was accompanied by an increased expression of Bax and Bak without consistent modulation of other Bcl-2 family proteins [21].

The apoptosis has been shown to be a major barrier to cancer cells [22]. Various factors such as Bcl-2 family members play a major role in the intrinsic mitochondrial apoptotic cascade [23]. The members of Bcl-2 family are divided into two main groups, pro-apoptotic (Bax, Bad, Bak, Bim, Bid, Bik, Noxa, Puma) and antiapoptotic (Bcl-2, Bcl-xL, Bcl-w, Mcl-1), which are characterized by sharing at least one region of Bcl-2 homology sequence termed BH1-BH4 [24, 25]. The over expression of survival proteins not only contributes to the progression of cancer, but also confers resistance to the therapeutic treatments [25]. Nowadays, the importance of apoptosis, its signalling pathways, checkpoints, mediators and modulators in the pathogenesis and prognosis of colorectal carcinoma is being increasingly recognized. Molecules involved in these pathways represent potential diagnostic markers and therapeutic targets and consequently are the focus of numerous research efforts [26]. Bax/Bcl-2 ratio can act as a rheostat which determines cell susceptibility to apoptosis as well as affects tumour progression and aggressiveness [27]. It has been reported that antiapoptotic Bcl-2 protein plays a role in colorectal precancerous and cancerous lesions [28]. On the other hand, the observed loss of Bcl-2 expression in high-grade tumours may lead to tumour progression to a deregulated state where Bcl-2 would not be required for the cell survival [29]. Indicating the involvement of Bax expression in tumour differentiation and metastatic progression, it has been demonstrated that Bax expression is decreased not only from primary to metastatic but also well/ moderately to poorly differentiated tumours, what leads to more infiltrative growth pattern, more distant metastases, and a weak trend toward poor prognosis [27]. The absence of Bax expression in colorectal cancer cells might induce resistance to apoptosis triggered by different chemotherapeutic agents [30]. The expression of Bcl-2 and Bax has been shown to predict the response to 5-FU based adjuvant therapy in colorectal carcinomas. Namely, the patients with low Bax/Bcl-2 ratio would benefit from this therapy [31]. Over 50 years have passed since 5-FU was developed, but it still plays a key role in chemotherapeutic regimens in the treatment of breast, colon and pancreatic cancer [32]. Although, 5-FU is a commonly used effective chemotherapeutic agent for treating a wide variety of malignant tumours, the effectiveness of the chemotherapy may be limited because of acquired or intrinsic drug resistance [33].
Small BH3-only members, with proapoptotic activity, may be helpful in elucidating alterations in Bax and Bcl-2 expression level in human colorectal carcinoma [27]. Proapoptotic and prosurvival Bcl-2 family proteins contain shared Bcl-2 homology domains BH1-3, that are involved in the formation of a hydrophobic BH3 docking groove harbouring binding sites for only BH3 containing proteins [34]. It has been demonstrated that peptides containing only BH3 domain of proapoptotic Bcl-2 family members are able to bind and inhibit antiapoptotic proteins [25]. Some BH3-only proteins such as Bim may directly bind to and activate Bax, what leads to the oligomerisation of this proapoptotic protein, permeabilization of the mitochondrial outer membrane, cytochrome c release and activation of caspase-9, -3, -6 and -7 [35, 36]. It has been suggested that Bim downregulation is important for tumour-genesis, especially for metastatic ability [36]. Recently, it has been shown that ALA upregulates Bim in ovarian carcinoma cells [9]. Bim phosphorylation by extracellular signal regulated kinase 1/2 (ERK1/2) promotes its degradation through the proteasome-ubiquitination system [36]. Some preclinical studies have pointed out that Bim induction by inhibition of the ERK pathway plays a key role in apoptosis of oncogene-addicted solid cancer cells including BRAF- mutant colorectal ones [37]. Moreover, it has been recently reported that BH3 mimic ABT-737 may markedly suppress the ERK1/2 phosphorylation levels in a concentration-dependent manner and induce proapoptotic Bim signalling pathway in human oral squamous carcinoma HN22 cells [38]. Contrary to Bim, the phosphorylation of Bcl-2 mediated by ERK2 might be of substantial importance for Bcl-2 stability and cell survival. Bcl-2 dephosphorylation induces its degradation through the proteasome-ubiquitination system and subsequent cell death [39]. It has been documented that ALA induces downregulation of Bcl-2 through its enhanced proteosomal degradation in human lung epithelial cancer cells [7]. The activation of ERK, c-Jun N-terminal kinase (JNK) and p38 kinase in colon tissues was shown to be significantly inhibited by ALA treatment [40]. Thus, by the inhibition of mitogen-activated protein kinase (MAPK)/ERK, ALA may be considered to upregulate Bim and downregulate Bcl-2 levels in cancer cells. Phosphorylation of Bcl-2 family members has appeared to be a generalized phenomenon that occurs in proapoptotic and antiapoptotic family members [41]. Bax conformational change and its translocation to the mitochondrial membrane is inhibited by phosphorylation in an Akt-dependent manner. After the phosphorylation, Bax effects on the mitochondria are inhibited by remaining its inactive heterodimerized form with antiapoptotic Bcl-2 family members in cytoplasm. Akt (protein kinase B, PKB), also, phosphorylates and inhibits the protease activity of caspase-9 [41-44].

It has been shown on hepatoma and human breast cancer cells that ALA may inhibit Akt activity [10, 11]. In that way, by the inhibition of Akt, ALA may potentiate Bax proapoptotic actions. Recent study on bladder cancer cells represents one more confirmation of the ERK1/2 and Akt downregulation caused by ALA [12]. Generally, the kinase activities of Akt and ERK1/2 were shown to be significantly upregulated in CD133+ primary colon cancer cells. The clonogenic growth, proliferation and differentiation of these cells was greatly reduced by the inhibition of Akt and ERK1/2 activity. The reported involvement of Akt and MAPK pathways in the tumorigenesis of CD133+ colon cancer cells, suggest that molecules in these two pathways might be potential targets in the future therapy [45]. Moreover, the fate of Caco-2 cells has been shown to be regulated by MAPK and Akt pathways [46, 47]. Thus, the potential mechanism of proapoptotic effect of ALA concretely in Caco-2 cells might be explained on the basis of these mentioned pathways.

The activation of intrinsic mitochondrial pathway of apoptosis in cancer cells, mediated by Bcl-2 family members as critical checkpoints, represents the most frequent mechanism of action of anticancer drugs [48]. Antiapoptotic family members are now a major target in the development of novel anticancer drug candidates [49]. Many structurally different Bcl-2 inhibitors and some Bax activators have been discovered in recent years [50, 51]. A large number of preclinical data suggest that BH3 mimetics may be highly useful for applying in synergistic therapies with conventional anticancer drugs and radiotherapy [35]. Despite this, the use of BH3 peptides as therapeutic agents may be limited by their unsatisfactory bioavailability, including poor cellular permeability, solubility and metabolic instability in vivo. It might be feasible to develop agents by "BH3 profiling" with individual prosurvival Bcl-2 family members like targets [52].

The search of protein targets of ALA as well as docking of the binding site is necessary to understand and support its physiological roles and pharmacological effects [2]. Considering the promising biological results in order to evaluate the mode of possible interaction of ALA and 5-FU with Bcl-2 and Bax targets, molecular docking of these compounds and some known Bcl-2 inhibitors and Bax activators was carried out and results were compared. The structures of known inhibitors/activators of these targets are various and were used as starting points to allow the evaluation of the binding modes of ALA and 5-FU. The binding modes of the two assayed compounds resembled those found by the docking study on complexes of Bcl-2 and Bax with their inhibitors and activators, respectively. Our docking study indicated that the binding mode of two BH3 mimetics with reported high affinity,
synthesized, orally available ABT-263 (navitoclax) and its synthetic derivative ABT-199, includes the interaction with Arg66 residue of Bcl-2 target. Namely, the interactions of the sulfonyl oxygen of ABT-263 and the carbonyl and sulfonyl oxygen and oxygen of the nitro group of highly specific and potent Bcl-2 inhibitor ABT-199 with Arg66 residue were established. Importantly, the interaction of carbonyl oxygen of ALA with Arg66 residue was found in the present study. The common interaction of the binding modes of ALA and 5-FU with Bcl-2 protein was shown to be the arene-hydrogen interaction with Tyr161, which was shown to be involved also in the binding of BH3 mimetic BI97D6, the new compound of gossypol family with modest binding affinity and reported in vivo antitumor activity. Most of the amino acid residues of the Bcl-2 binding site for ALA and 5-FU overlapped with those involved in the binding pockets for the known and assayed Bcl-2 inhibitors (Figure 2).

Carbonyl oxygen of ALA was found to be responsible also for the binding of Bax target. Namely, the hydrogen bond interaction between carbonyl oxygen of ALA and 5-FU with the backbone structure of Asp53 was found to be involved in their binding for this pro-apoptotic target. The amino acid residues of the Bax binding site for ALA and 5-FU mainly overlapped with those of the binding cavity for Bax activators BAM-7 and BTC-8, but the arene-hydrogen interactions involved in the binding mode of BAM-7 and BTC-8, were found absent in the binding of ALA and 5-FU (Figure 3).

Molecular docking was performed to evaluate the probable mechanism of proapoptotic action of ALA in cancer cells. It was found that some interactions with the certain amino acid residues of Bcl-2 and Bax binding sites might play a role in the mechanism of Bcl-2 downregulation and Bax upregulation caused by ALA. Moreover, the binding modes of ALA and 5-FU with both targets were shown to be closely similar, and some interactions the same like those of standard known BH3 mimetics. Therefore, this may be a new possible mechanism of the proapoptotic effect of these compounds.
In order to open new possibilities of this targeted medical application of ALA and to determine the potential advantages with respect to existing agents that influence the apoptotic markers and pathways in cancer cells, the calculated physico-chemical properties of this compound were taken into account. Summarizing the physico-chemical properties, calculated by using Molinspiration tool [16], it was concluded that this compound obeys the Lipinski "Rule of five" and meets all criteria for good solubility, permeability and conformational flexibility. More than one violation of this rule is the critical limit for acceptable drug-likeness [53]. It might be concluded that ALA also obey the Veber rules [54] and might be considered to possess the ability to penetrate biological membranes, that is a common requirement for bioavailability. The lipophilicity of compounds is a common property used to estimate the membrane permeability of a molecule. Thus, ALA may easily cross biological membranes, and reach all the compartments of the cell. Therefore, this is one more confirmation that ALA would be easily delivered to the targets inside the cell or on the cell membranes.

Transcription nuclear factor kappa B (NF-κB) is present in the cytoplasm and translocates into the nucleus in response to various inflammatory stimuli. It is known that NF-κB regulates the expression of genes which play a role in the development and progression of cancer such as cell proliferation, migration and apoptosis [55]. Expression of inducible transcription factor NF-κB and mitochondrial markers may be helpful in predicting clinical outcome and good predictor of cellular response to a given chemotherapeutic agent. As indicated in our previous studies, the inhibition of growth of cancer HeLa and Caco-2 cell lines associated with the inhibition of NF-κB, this powerful transcription factor in the regulation of cell fate might also be one of the potential ways through which an antiproliferative effect of ALA is performed [8].

To determine the chemo-preventive potential and prognostic relevance of biomarkers involved in the apoptotic pathways in colorectal cancer, multiple markers that reflect the apoptotic status should be studied together. According to our results, ALA may be considered as a promising therapeutic agent in colon cancer due to its efficiency and significant chemopreventive potential. An ideal therapeutic agent would specifically target the cancer cells without causing serious cytotoxicity and systemic damage.
However, in the case of conventional chemotherapeutic agents, the reason of serious adverse effects is the presence of their targets in normal healthy cells. High selective or specific actions with high efficacy are needed for a good drug candidate that would be nontoxic to the patient [38]. The advantage of ALA from this point of view is the evidence that it may inhibit proliferation and induce apoptosis in various cancer cell lines without effect on normal non-transformed cells [7].

Conclusions

Our findings suggest that the possible mechanism of proapoptotic effect of ALA in human colon cancer cells may be the inhibition of Bel-2 or the activation of Bax apoptotic checkpoints and regulators. Thus, ALA may be considered as potential BH3 mimetic. Moreover, the possible mechanism may involve the inhibition of ERK1/2 and Akt pathways. The supposed mechanisms may act synergistic in exerting pro-apoptotic effects of ALA in cancer cells. Considering the physico-chemical properties taken into account, it may be concluded that ALA can be easily delivered to the potential intracellular targets, such as Bel-2, Bax, ERK1/2, Akt or NF-κB. These findings may be a part of the explanation of possible mechanisms that contribute to the beneficial effects of this readily available dietary supplement in cancer therapy. The observed mechanisms represent a step forward that will be helpful to further investigation. The obtained results are only the beginning of the examination of the possible effects of ALA on cancer chemoprevention and its effects on standard cytostatics. Moreover, further research from different aspects may be useful in the way of clarification of these suggested or some new potential mechanisms of ALA-mediated antiproliferative and proapoptotic effects in the various types of cancer cells.

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